# I. Taxonomy, Fermentation, Isolation and Biological Activities

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Selective growth inhibition against IL-6 dependent cells was detected in fermentation extracts of a fungal strain FO-4649 which was characterized as *Sporothrix* species. An active metabolite (1) termed chlovalicin was isolated together with ovalicin and two other ovalicin derivatives (compounds **3** and **4**). Chlovalicin, a ovalicin derivative with a chlorinated methylene moiety at the C-1 position of the cyclohexane ring, dose-dependently inhibited the growth of IL-6 dependent MH60 cells (IC<sub>50</sub>, 7.5  $\mu$ M) in the presence of 0.2 U/ml IL-6 and, to a lesser extent, the growth of B16 melanoma cells (IC<sub>50</sub>, 38  $\mu$ M). Among the other three compounds, only ovalicin showed inhibitory activity (IC<sub>50</sub>, 27  $\mu$ M) against MH60 cells. These four compounds did not show any antimicrobial activity at a concentration of 1000  $\mu$ g/ml.

In the course of a screening program for new types of antibiotics showing cytocidal activity, four structurally related compounds were discovered from the fermentation broth of *Sporothrix* sp. FO-4649, which was isolated from a soil sample. One appeared to be a new compound and was designated as chlovalicin (1), and the other compounds were identified as ovalicin (2) and its derivatives  $(3, 4)^{1,2}$  (Fig. 1). The present paper outlines taxonomic studies of the producing strain, and the production, isolation and biological activity of the new compound, chlovalicin.

### **Materials and Methods**

#### **General Experimental Procedures**

The fungal strain FO-4649, isolated from a soil sample, was used for production of chlovalicin (1). DC-Alufolien Kieselgel  $60F_{254}$  (Merck) was used for TLC analysis. HPLC was carried out using the HITACHI D-2000 system and an ODS packed column (Senshu Pak Pegasil ODS,  $5 \,\mu$ m, i.d.  $20 \times 250$  mm).

## Taxonomic Studies

The fungal strain FO-4649 was originally isolated from a soil sample. For identification of the fungi, potato-dextrose agar, malt extract agar, CZAPEK's agar and LCA (MIURA's medium) agar were used. Morphology was observed by microscopy (Olympus Vanox-S AH-2).

# Antimicrobial Activity

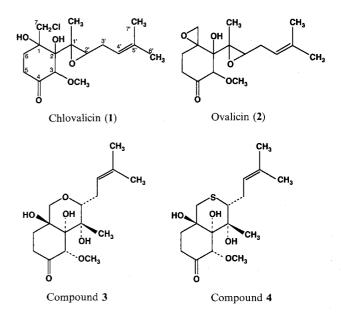
Antimicrobial activity was tested using paper disks (i.d. 6mm, ADVANTEC). Bacteria were grown on

Mueller-Hinton agar medium (Difco), and fungi and yeasts were grown on potato broth agar medium. Antimicrobial activity was observed after 24-hour incubation at  $37^{\circ}$ C for bacteria and after 48-hour incubation at  $27^{\circ}$ C for fungi.

#### Cell Lines

P388/S (mouse leukemia), P388/ADM (adriamycin resistant cell line), L929 (mouse fibroblast), HeLa (human cerical carcinoma), and B16 (mouse melanoma)

Fig. 1. Structures of chlovalicin (1), ovalicin (2) and related compounds (3, 4).



cell lines were kindly provided by Dr. INABA, Cancer Research Institute. IL-6 dependent MH60 cells were kindly provided by Prof. OH-ISHI, Kitasato University.

## Cytotoxic Activity Tests

Five cell lines were maintained in monolayers or in suspension in EAGLE's minimum essential medium (MEM) supplemented with 10% fetal calf serum (FCS) or RPMI 1640 medium supplemented with 10% FCS, respectively. To determine the cytotoxicity of chlovalicin, cells suspended in 200  $\mu$ l of the medium (1~2×10<sup>4</sup> cells/ml) were plated in a 96-well culture plate (Corning) and incubated for 24 hours at 37°C in a 5% CO<sub>2</sub>-95% air atmosphere. To each well was added 5  $\mu$ l of medium containing different concentrations of chlovalicin. After 72 hour incubation, the cell growth was measured colorimetrically by the tetrazolium salt (MTT) method<sup>3)</sup>.

MH60 cells ( $5 \times 10^3$  cells) suspended in 100  $\mu$ l of RPMI 1640 medium supplemented with 10% FCS were plated in a 96-well culture plate and incubated with  $5 \mu$ l of test samples for 72 hours at 37°C in the presence of 0.02 U rhIL-6 (100  $\mu$ l) in a 5% CO<sub>2</sub>-95% air atmosphere. The cell growth was evaluated by the MTT method.

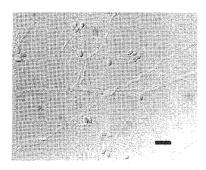
### **Results and Discussion**

### Taxonomy of Producing Organism

The fungal strain FO-4649 was isolated from a soil sample. For identification of the fungi, potato-dextrose agar, malt-extract agar, CZAPEK's agar and LCA (MIURA's medium) agar were used. This strain grew moderately to form oyster white to light brownish gray colonies with a diameter of  $20 \sim 40$  mm after inoculation for 7 days at 25°C. The colony surface was floccose on various agar media. The reverse color was pale yellow to grayish white. Morphological observation was done under a microscope (Olympus Vanox-S AH-2). When the strain FO-4649 was grown on potato-dextrose agar at 25°C for 7 days, the conidia were formed by blastoconidia from direct substrate hyphae and sympodioconidia

Fig. 2. Photomicrograph of strain FO-4649 on potatodextrose agar.

Bar represents 20 µm.



from conidiophores. Conidia were obovoid or obpyriform, smooth, hyaline to slightly colored, unicellular,  $5 \sim 12 \times 3 \sim 6 \,\mu\text{m}$  in diameter. The temperature permitting growth of the strain was 12 to 35°C, and optimum temperature for growth was 15 to 30°C. A light micrograph of this strain is shown in Fig. 2.

From the above characteristics, strain FO-4649 was identified as a member of the genus *Sporothrix*<sup>4,5)</sup>, and named *Sporothrix* sp. FO-4649. It was deposited at the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology Japan, as FERM P-14796.

## Fermentation

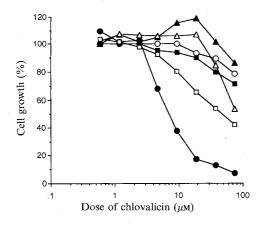
The stock culture of the producing organism was inoculated into a test tube  $(20 \times 200 \text{ mm})$  containing 10 ml of a seed medium [glucose 2%, yeast extract 0.2%, polypeptone 0.5%, MgSO<sub>4</sub>·7H<sub>2</sub>O 0.05%, KH<sub>2</sub>PO<sub>4</sub> 0.1% and agar 0.1% (adjusted to pH 6.0 before sterilization)]. The test tube was incubated at 27°C for 72 hours on a reciprocal shaker. The resulting culture  $(20 \times 10 \text{ ml})$  was transferred to 100 ml of production medium  $(100 \times 100 \text{ ml})$  consisting of glucose 1.0%, tryptone 0.5%, yeast extract 0.3%, maltose 0.3%, and agar 0.1%, (adjusted to pH 6.5 before sterilization) in a 500-ml Erlenmeyer flasks, and the fermentation was carried out at 27°C for 6 days with an agitation rate of 200 rpm. Detection of the chlovalicin in the fermentation broth was followed by growth inhibition on the IL-6 responsive cell system. After 6 days of fermentation, the amount of chlovalicin in the broth filtrate reached a maximum (0.35  $\mu$ g/ml).

# Extraction and Isolation of Active Compounds

The culture broth (10 liters) of *Sporothrix* sp. FO-4649 was filtered, the resulting supernatant was extracted with EtOAc (10 liters  $\times$  2) and then the organic layer was dried with Na<sub>2</sub>SO<sub>4</sub> anhydride. The combined EtOAc layers were concentrated under reduced pressure to give a brown oil (1.75 g). The oily residue was applied to silica gel column chromatography (E. Merck, Kieselgel 60, 70 ~ 230 mesh). The materials were eluted with a step wise gradient of CHCl<sub>3</sub>-CH<sub>3</sub>OH (100:0, 100:1, 50:1, 20:1, 10:1, each 500 ml). Active fractions (CHCl<sub>3</sub>-CH<sub>3</sub>OH, 50:1) were concentrated *in vacuo* to yield an oily material (100 mg).

Further purification of chlovalicin was carried out by HPLC using a preparative ODS column (PEGASIL ODS (i.d.  $20 \times 250$  mm); Senshu Science Co. LTD., Japan; solvent: 60% aq CH<sub>3</sub>CN: UV 210 nm, flow rate 7 ml/minute) to give a colorless oil (30.2 mg) (mixture of Fig. 3. Effect of chlovalicin (1) on the growth of various mammalian cells.

•: MH60 (IL-6 dependent), ▲: P388C, △: P388/ADM, ○: L929, ■: HeLa, □: B16.



chlovalicin and its derivatives). Final purification was carried out by HPLC using a preparative ODS column (PEGASIL ODS H-5251 (i.d.  $20 \times 250$  mm); Senshu Science Co. LTD., Japan; solvent: 50% aq CH<sub>3</sub>CN: UV 210 nm, flow rate 7 ml/minute). Pure chlovalicin (1) (3.5 mg), ovalicin (2) (12.3 mg), compound 3 (2.5 mg) and compound 4 (2.6 mg) were obtained.

Structures of chlovalicin and the related compounds are shown in Fig. 1. Studies on the structural determination of chlovalicin will be reported in a separate paper.

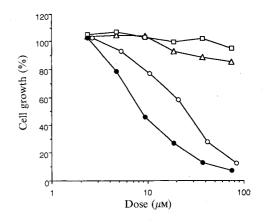
## **Biological Activity**

Growth inhibitory activity of chlovalicin was examined using various mammalian cells *in vitro* (Fig. 3). The antibiotic markedly inhibited growth of IL-6 dependent MH60 cells in a dose dependent manner. The IC<sub>50</sub> values for MH60 and B16 melanoma were 5.7  $\mu$ M and 38  $\mu$ M, respectively. While the IC<sub>50</sub> values for P388/S leukemia, P388 leukemia/ADM, HeLa S3 carcinoma and L929 fibroblast were all more than 75  $\mu$ M.

The cytotoxicities of chlovalicin, ovalicin and other related compounds against IL-6 dependent MH60 cells were further examined. MH60 cells were incubated with chlovalicin, ovalicin, compound **3**, or compound **4** for 72 hours in the presence of 0.2 U/ml of IL-6. As shown in Fig. 4, chlovalicin showed higher activity about 3 times of ovalicin (IC<sub>50</sub>, 27  $\mu$ M) but compound **3** and compound **4** were not active.

These four antibiotics showed no antimicrobial activities at a concentration of  $1000 \,\mu\text{g/ml}$  against Staphylococcus aureus KB34 (FDA 209p), Micrococcus luteus KB40 (PCI 1001), Bacillus subtilis KB27 (PCI 219), Mycobacterium smegmatis KB46 (ATCC 607),

- Fig. 4. Effect of chlovalicin and other compounds on the growth of IL-6-dependent MH60 cells.
- •: Chlovalicin (1),  $\bigcirc$ : ovalicin (2),  $\triangle$ : compound 3,  $\Box$ : compound 4.



Escherichia coli KB 8 (NIHJ), Pseudomonas aeruginosa KB 105 (P-3), Xanthomonas oryzae KB 88, Candida albicans KF1, Saccharomyces sake KF26, Aspergillus niger KF 103 (ATCC-6725), Piricularia oryzae KF 180, Mucor racemosus KF 223, Clostridium perfringens KB 129, Bacteroides fragilis KB 169, and Acholeplasma laidlawii PG-8.

Chlovalicin dose-dependently inhibited the growth of IL-6 dependent MH60 cells (IC<sub>50</sub>, 7.5  $\mu$ M) in the presence of IL-6. MH60 cells seemed the most sensitive among these cells lines, so it is not clear whether chlovalicin selectively inhibits the IL-6 activity. Further examinations are necessary to clarify the mechanism of chlovalicin and the specificity for other cytokines.

It has been suggested that abnormal IL-6 production is closely related to the progression of cancer cachexia<sup>6~8)</sup> and hormone-dependent hypercalcemia<sup>9,10)</sup>, and to the development of multiple myeloma<sup>11,12)</sup> and rheumatoid arthritis<sup>13,14)</sup>. Only a few substances are known to suppress IL-6 activity, *i.e.*, suramin<sup>15)</sup> and monoclonal antibodies<sup>16)</sup>. Therefore, IL-6 inhibitors will be useful in studying its role in disease.

Besides our present report concerning the inhibitory effect of chlovalicin on IL-6-dependent cell growth, recently, a new activity of chlovalicin as IgE production inhibitor was reported<sup>17)</sup>. Therefore, chlovalicin is expected to be a new type anti-allergic drug possessing the inhibitory effects to both IgE production and IL-6 activities.

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#### References

- BOLLINGER, P.; H.-P. SIGG & H.-P. WEBER: Die struktur von ovalicin. Helvetica Chimica Acta. 56: 819~830, 1973
- HARTMANN, G. R.; H. RICHTER, E. M. WEINER & W. ZIMMERMANN: On the mechanism of action of the cytostatic drug anguidine and of the immunosuppressive agent ovalicin, two sesquiterpenes from Fungi. Planta Medica 34: 231~252, 1978
- 3) ALLEY, M. C.; D. A. SCUDIERO, A. MONKS, M. L. HURSEY, M. J. CZERWINSKI, D. L. FINE, B. J. ABBOTT, J. G. MAYO, R. H. SHOEMAKER & M. R. BOYD: Feasibility of drug screening with panel of human tumor cell lines using a microculture tetrazolium assay. Cancer Res. 48: 589~ 601, 1988
- BARRON, G. L.: The genera of hyphomycetes from soil. The Williams & Wilkins Co.: 284~285, 1968
- 5) ARX, J. A. V.: The genera of fungi-sporulating in pure culture (3rd. ed.) J. Cramer, Vaduz.: 283~331, 1981
- LANGSTEIN, H. N. & J. A. NORTON: Mechanisms of cancer cachexia. Hematol. Oncol. Clin. North Am.: 103~107, 1991
- NORTON, J. A.; J. L. PEACOCK & S. D. MORRISON: Cancer cachexia. CRC Crit. Rev. Oncol. Hematol. 7: 289~327, 1987
- STRASSMANN, G.; M. FONG, J. S. KENNEY & C. O. JACOB: Evidence for the involvement of IL-6 in experimental cancer cachexia. J. Clin. Invest. 89: 1681~1684, 1992
- 9) WEISSGLAS, M.; D. SCHAMHART, C. LOWIK, S. PAPAPOU-LOS, P. VOS & K. H. KURTH: Hypercalcemia and cosecretion of interleukin-6 and parathyroid hormone related peptide by a human renal cell carcinoma im-

planted into nude mice. J. Urol. 153: 854~857, 1995

- 10) VANDERSCHUEREN, B.; J. C. DUMON, V. OLEFFE, C. HEYMANS, J. GERAIN & J. J. BODY: Circulating concentrations of interleukin-6 in cancer patients and their pathogenic role in tumor-induced hypercalcemia. Cancer Immunol. Immunother. 39: 286~290, 1994
- 11) KLEIN, B.; X. G. ZHANG, M. JOURDAN, J. M. BOIRON, M. PORTIER, Z. Y. LU, J. WIJDENES, J. BROCHIER & R. BATAILLE: Interleukin-6 is the central tumor growth factor *in vitro* and *in vivo* in multiple myeloma. Eur. Cytokine Netw. 1: 193~201, 1990
- FATTORI, E.; C. DELLA-ROCCA, P. COSTA, M. GIORGIO, B. DENTE, L. POZZI & G. CILIBERTO: Development of progressive kidney damage and myeloma kidney in interleukin-6 transgenic mice. Blood. 83: 2570~2579, 1994
- 13) KAISHO, T.; K. ORITANI, J. ISHIKAWA, M. TANABE, O. MURAOKA, T. OCHI & T. HIRANO: Human bone marrow stromal cell lines from myeloma and rheumatoid arthritis that can support murine pre-B cell growth. J. Immunol. 149: 4088 ~ 4095, 1992
- MONTERO-JULIAN, F. A.; B. KLEIN, E. GAUTHEROT & H. BRAILLY: Pharmacokinetic study of anti-interleukin-6 (IL-6) therapy with monoclonal antibodies: enhancement of IL-6 clearance by cocktails of anti-IL-6 antibodies. Blood. 85: 917~924, 1995
- 15) STRASSMANN, G.; M. FONG, C. E. FRETER, S. WINDSOR, F. D'ALESSANDRO & R. P. NORDAN: Suramin interferes with interleukin-6 receptor binding *in vitro* and inhibits colon-26-mediated experimental cancer cachexia *in vivo*. J. Clin. Invest. 92: 2152~2159, 1993
- 16) BRAKENHOFF, J. P.; F. D. DE-HON, V. FONTAINE, E. TEN-BOEKEL, H. SCHOOLTINK, S. ROSE-JOHN, P. C. HEINRICH, J. CONTENT & L. A. ARDEN: Development of a human interleukin-6 receptor antagonist. J. Biol. Chem. 269: 86~93, 1994
- ICHITOU, K.; K. MAGOME & T. YAGUCHI (Meiji seika kaisya, Ltd.): New bioactive compound, PF1131. Jpn. Kokai 17957 ('95), Jan. 20, 1995